

### National Wastewater Surveillance Programme - COVID-19

Weeks 11 & 12 (Weeks ending 19 March & 26 March 2023)

Report prepared on 29 March 2023

#### 99%

sites tested had SARS-CoV-2 detected (85/86 sites)

#### 70%

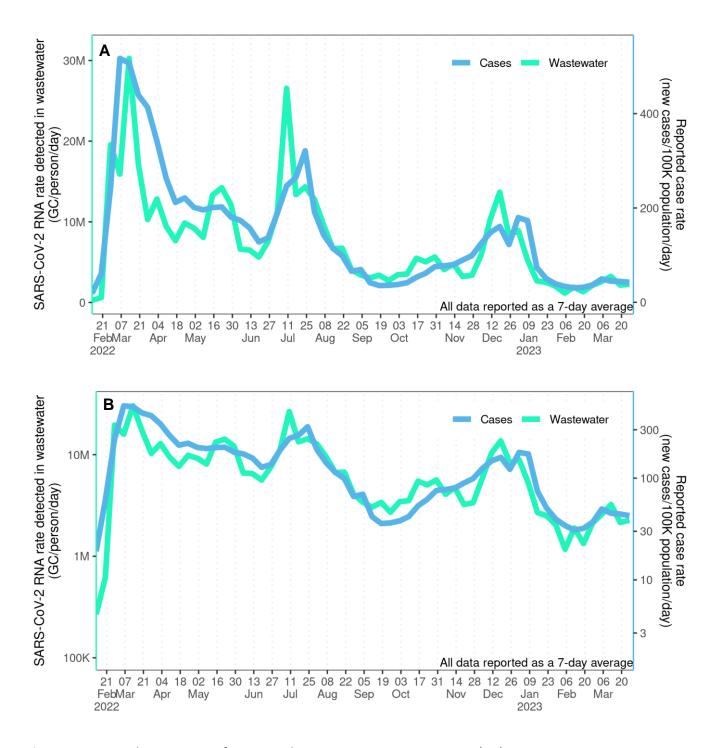
NZ population covered by wastewater testing past fortnight

# Omicron XBB (~64%)

Most prevalent variant detected in week 12

## Nationally, SARS-CoV-2 levels remain low and steady. Variant analysis suggests that XBB and CH.1.1 continue to be the most common variants detected.

- Comparing week ending 26 March to week ending 19 March 2023, 39% of sites show an increase in SARS-CoV-2 levels while 27% sites showed a decrease in SARS-CoV-2.
- Comparing week ending 26 March 2023 to one month ago (week ending 26 February 2023), 37% of sites show and increase in SARS-CoV-2 levels while 29% of sites showed a decrease in SARS-CoV-2 levels.
- The main variants detected in wastewater in the week ending 26 March 2023 (week 12) were XBB (includes XBB.1.5, ~64%) and CH.1.1 (~24%). Minor contributions from BA.2.75\* (includes XBF, ~8%), BQ.1.1 (~3%) and BA.2 (~1%). XBC and BA.4/BA.5 not detected.
- While many sites have recommenced their regular sampling following weather-related impacts in late January and February 2023, sampling from Napier is not possible.



**Figure 1.** National timeseries of estimated SARS-CoV-2 genome copies (GC) in wastewater rate (GC/person/day, green line) and reported case rate (new cases/100,000 population/day, blue line) on a linear scale (**A**) and Log<sub>10</sub> scale (**B**). Data reported as 7-day average.

### Results for Weeks 11 & 12 (Weeks ending 19 March & 26 March 2023)

In the two weeks ending 26 March 2023, 230 samples were collected from 86 locations across New Zealand. SARS-CoV-2 RNA was detected in 226/230 samples from 85/86 (99%) sites (Figure 2, Table 1).

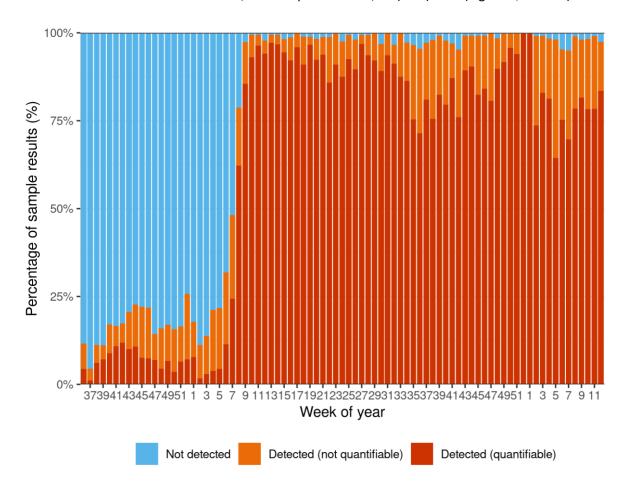
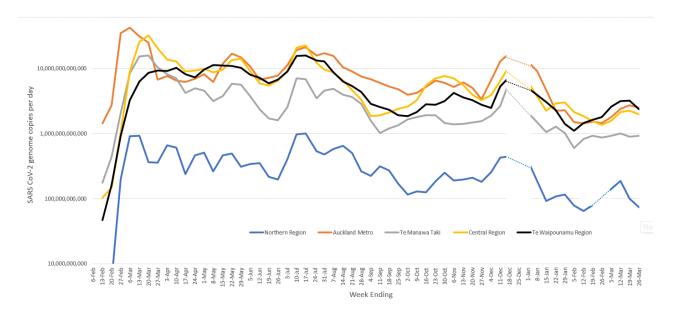


Figure 2. Results for SARS-CoV-2 RNA in wastewater collected across New Zealand.

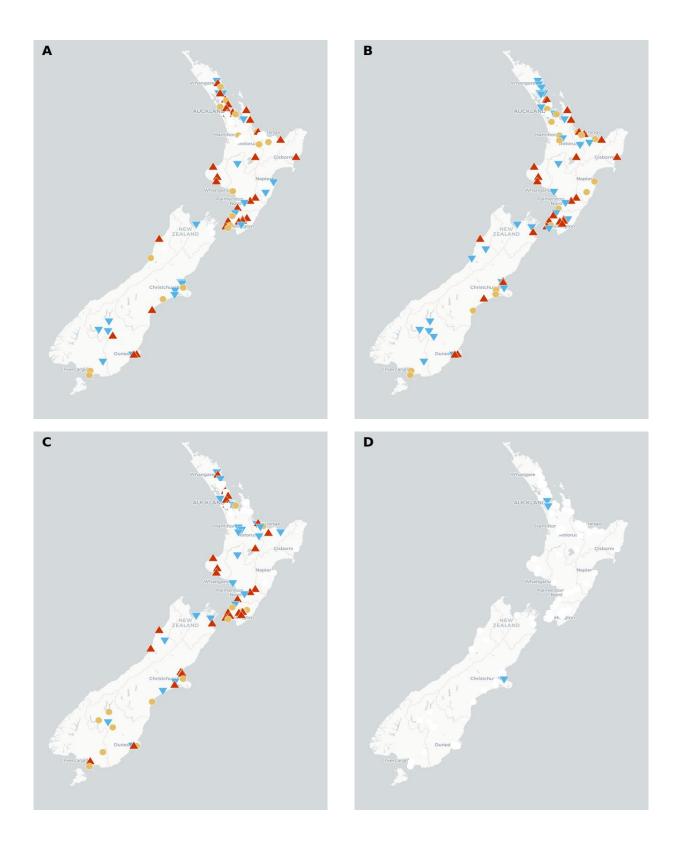
#### **Regional Trends**

Regional summaries (Figure 3) of the wastewater data indicates generally steady viral levels in all regions in weeks 11 and 12 compared to previous weeks. Note that regional trend analysis for week 52 (2022) and week 1 (2023) was only possible for Auckland Metro, as there were limited samples collected during the holiday period. Viral quantitation for the other regions were therefore not available during this period (denoted by dashed line).

Due to the weather-related impacts in February 2023, fewer samples were collected in some regions. The Central regional summary excludes Hawkes Bay samples in weeks 8-10, and analysis for the Northern region was not possible in between weeks 7-9 due to too few samples being received from this region. The dashed blue line shows inferred level in this region during this time (Figure 3).



**Figure 3.** Total SARS-CoV-2 genome copies detected per day in the five Ministry of Health regions. Dashed lines are inferred levels during periods when samples were either not collected (Christmas period) or insufficient numbers collected (due to weather impacts) for the region.



**Figure 4.** Comparison of SARS-CoV-2 levels for the week ending 12 March 2023, compared to levels measured: A) 1 week ago; B) 2 weeks ago; C) 4 weeks ago; D) 12 weeks ago. Only sites with results for both time points are included. When the viral quantity is 30% or more higher this is labelled as increased (red up arrow on map). When the viral quantity is 30% or more lower, this is labelled as decreased (blue down arrow on map). If viral levels have changed less than this in the compared weeks, this is labelled as no change (yellow circle on map). Interactive map of weekly results available publicly at https://www.poops.nz/

#### **Wastewater Variant Analysis**

In collaboration with Wilderlab, ESR generated the variant analysis results (Table 1, Figure 5) from sentinel sites in week 11 (ending 19 March 2023) and week 12 (ending 26 March 2023).

Wastewater variant analysis is based on sequencing a short fragment of the spike gene and therefore provides less resolution than WGS from clinical cases. As such, some specific lineages cannot be distinguished from each other, and are reported as variant groups. The following variants/groups are reported: BA.4/BA.5, BA2.75\* (includes BA.2.75/XBF/BR.2 subvariants), CH.1.1, BQ.1.1, XBB (includes XBB.1.5) and XBC.

Due to the increasing complexity of variants in the population, each at relatively low levels, the current approach for sequencing wastewater samples needs to be more precise to report percentages for each variant at the sentinel site level. Instead, the presence of each lineage will currently be reported. ESR is actively testing and developing methods to address the current uncertainty and increase the resolution to identify variants in wastewater.

Consistent with the WGS of clinical cases, the CH.1.1 subvariant will now be reported separately from other BA.2.75\* subvariants.

#### Results for weeks 11 and 12

Due to generally low viral levels in wastewater samples this fortnight, many sites did not return a reliable sequencing result. In weeks 10 and 11, 11/19 and 14/19 sites respectively provided a sequencing result. Results below (Table 1, Figure 5) relate to those sites that did produce reads that mapped to variants.

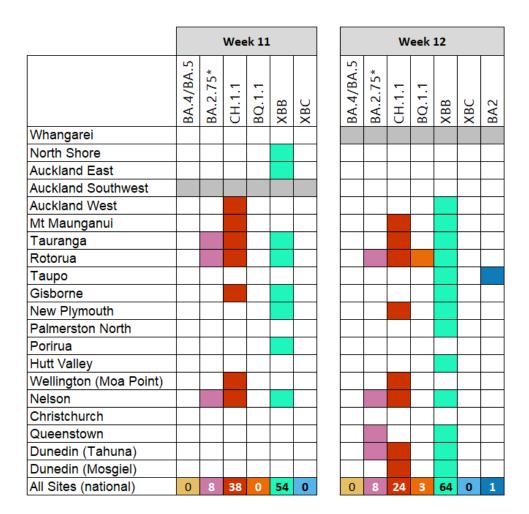
**XBB (includes XBB.1.5)** was **frequently detected**, comprising  $^{\sim}54\%$  of reads nationally in week 11 (8/11 sites), and  $^{\sim}64\%$  of reads nationally in week 12 (13/14 sites).

**CH.1.1** was also **frequently detected** in weeks 11 (7/11 sites) and 12 (8/14 sites). CH.1.1 comprised ~38% of sequencing reads nationally in week 11, declining to ~24% sequencing reads in week 12. Other subvariants in the BA.2.75\* group (including BM.4, BR.2, XBF and BA.2.75) accounted for another ~8% reads in both weeks.

BQ.1.1 was not detected in week 11 and accounted for only ~3% of national sequence reads in week 12, being detected in Rotorua only.

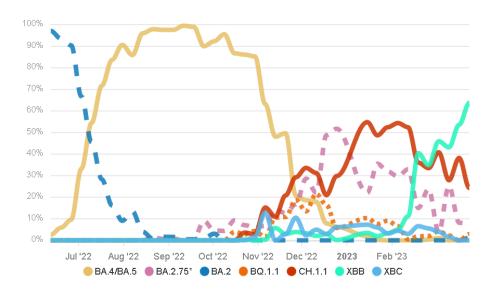
Neither the XBC nor BA.4/BA.5 variants were detected in either week.

For the first time in many weeks, the BA.2 variant was detected, at one site (Taupo) in week 12, comprising 1% of national sequencing reads that week.



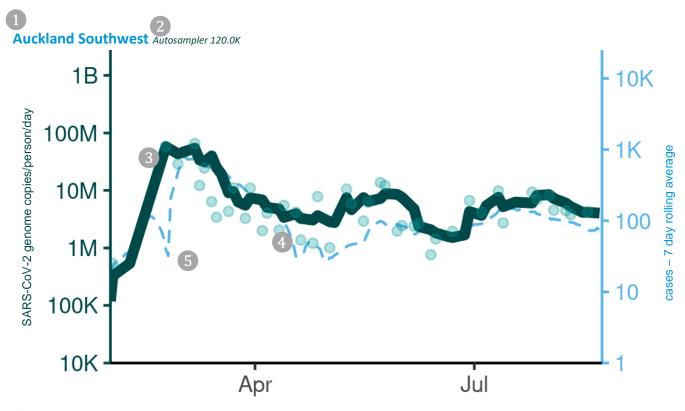
**Table 1**. Data from 19 wastewater sentinel sites sampled in week 11 (ending 19 March 2023) and week 12 (ending 26 March 2023) using a S-gene (spike) barcoding assay able to 'call' the BA.4/BA.5, the BA2.75\* constellation (includes BA.2.75/XBF/BR.2 subvariants), CH.1.1, BQ.1.1, XBB (includes XBB.1.5) and XBC (sub)variants. Coloured box denotes that the variant was detected at that site that week, white box denotes that the variant was not detected, and grey box denotes site was not sampled that week. Numbers in the bottom row denote the estimated percentage of each variant at the national scale.

#### **Variant Timeline - National**



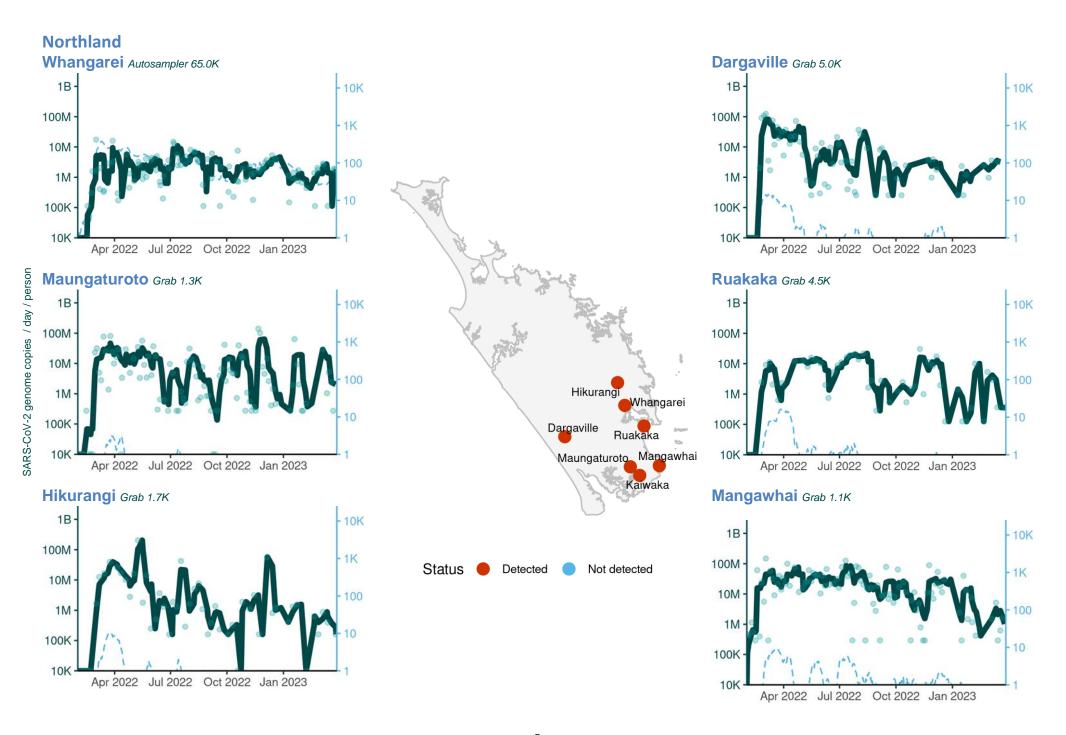
**Figure 5.** Change in variant prevalence over time at a national scale. Data are collected from up to 20 sentinel sites each week.

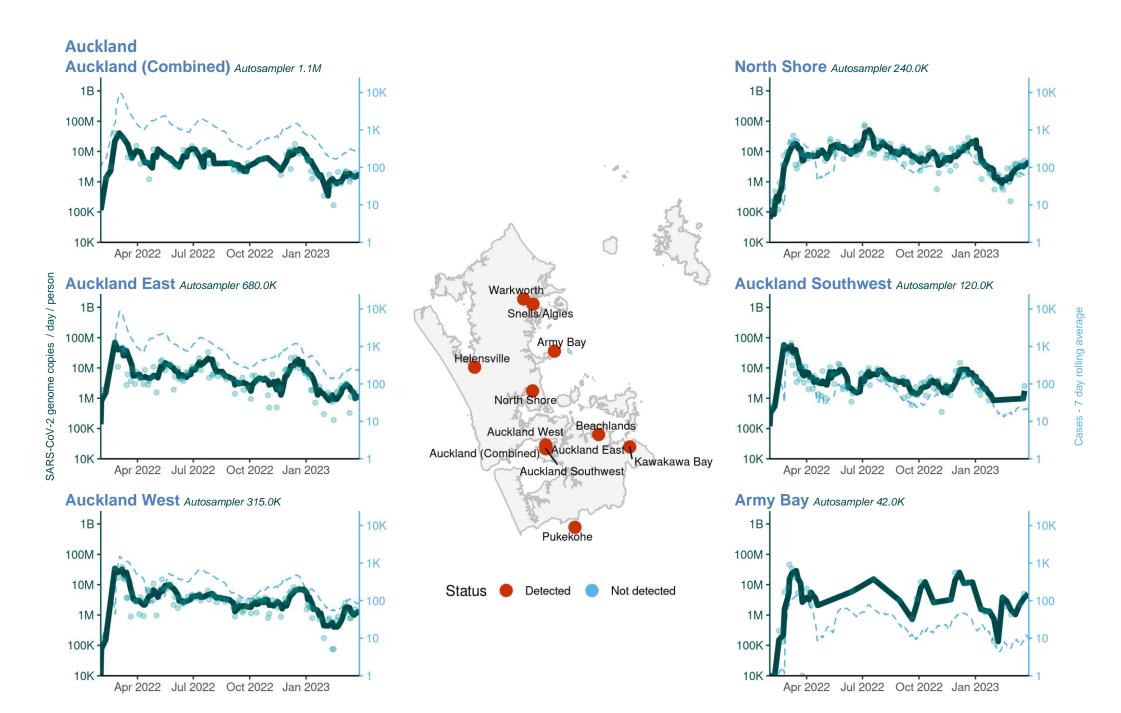
### **Interpreting Sites Graphs**

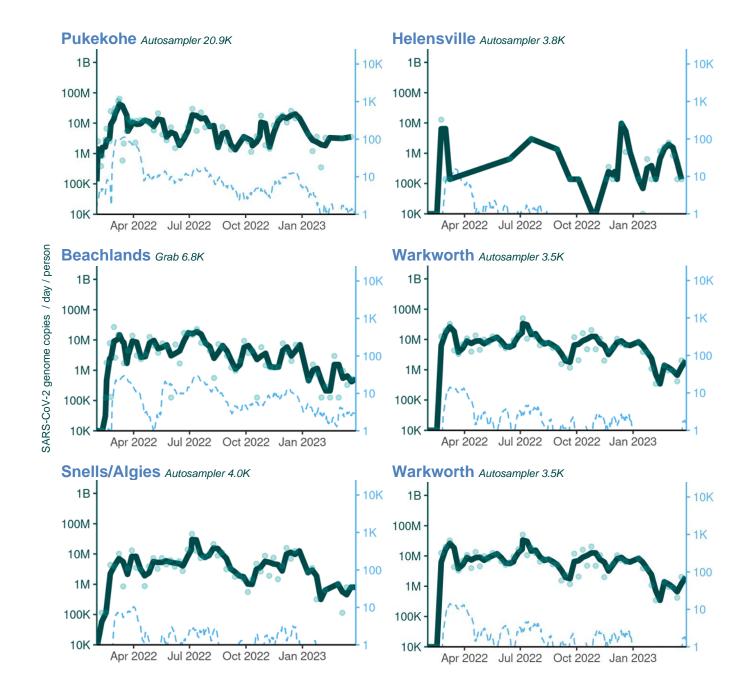


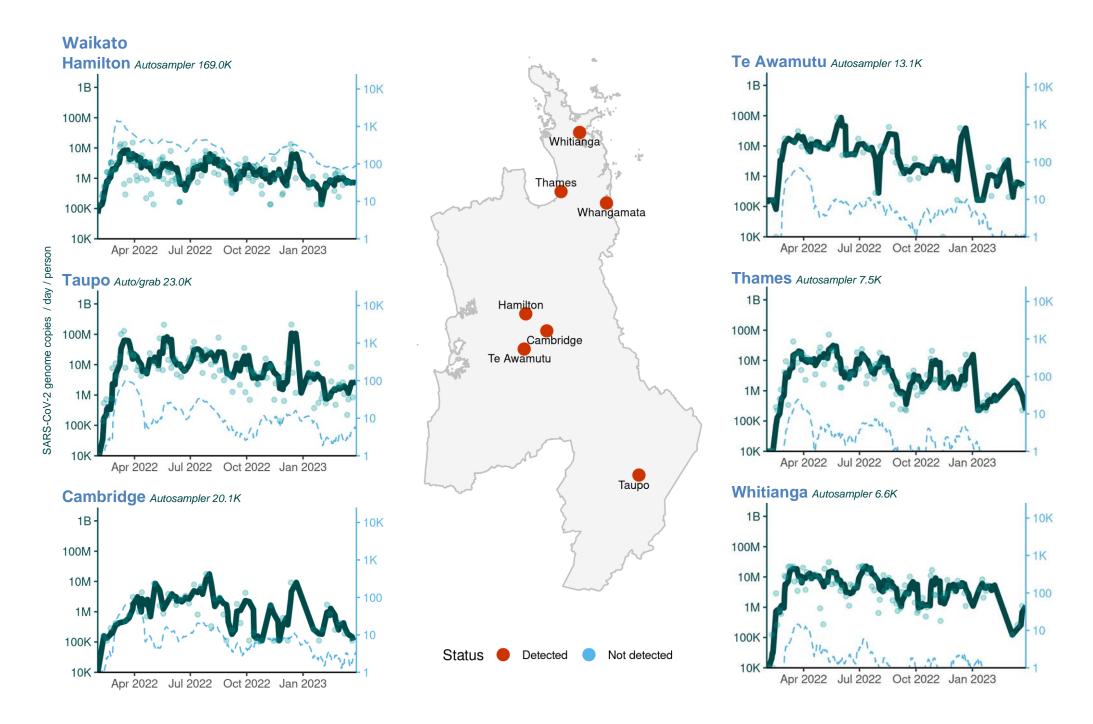
- Site name
- 2 Sample collection method and population. Results based on autosampler may be more representative than grab sample-based results.
- 3 Wastewater results shown as solid line | 14-day average of genome copies/person/day on a  $log_{10}$  scale.
- Individual results samples shown as circles | Rolling 14-day average of genome copies/person/day on a log<sub>10</sub> scale.
- 6 Rolling 7-day average of new cases shown as dashed line | New cases reported in a catchment based on reported date of illness on a log<sub>10</sub> scale. This data is not available for all sites and subject to change.

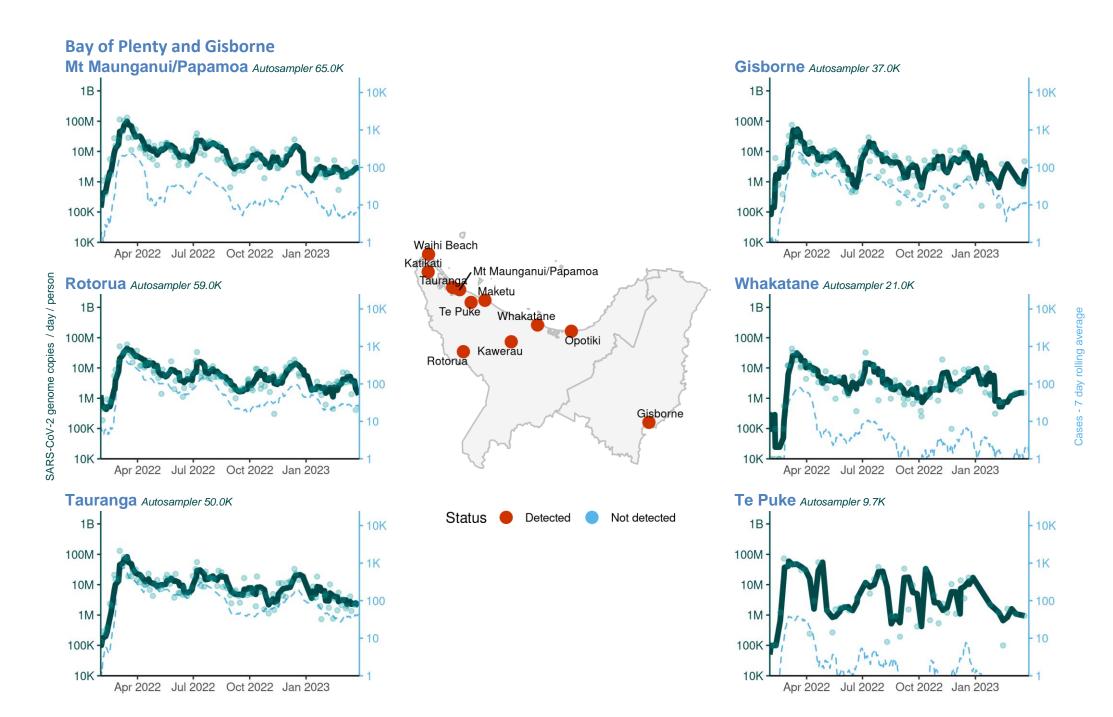
Note: Wastewater and cases data are on a log<sub>10</sub> scale. Scales on all graphs have been normalized to cover the same scale on every graph. Care should be taken when interpreting the data.



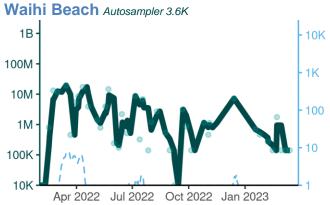


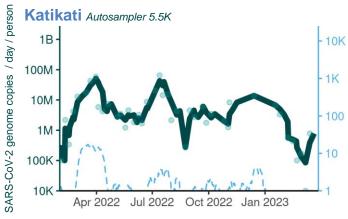


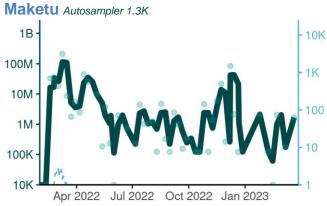


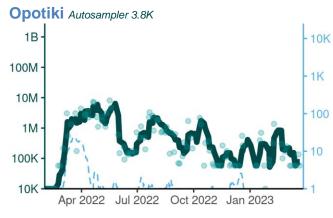


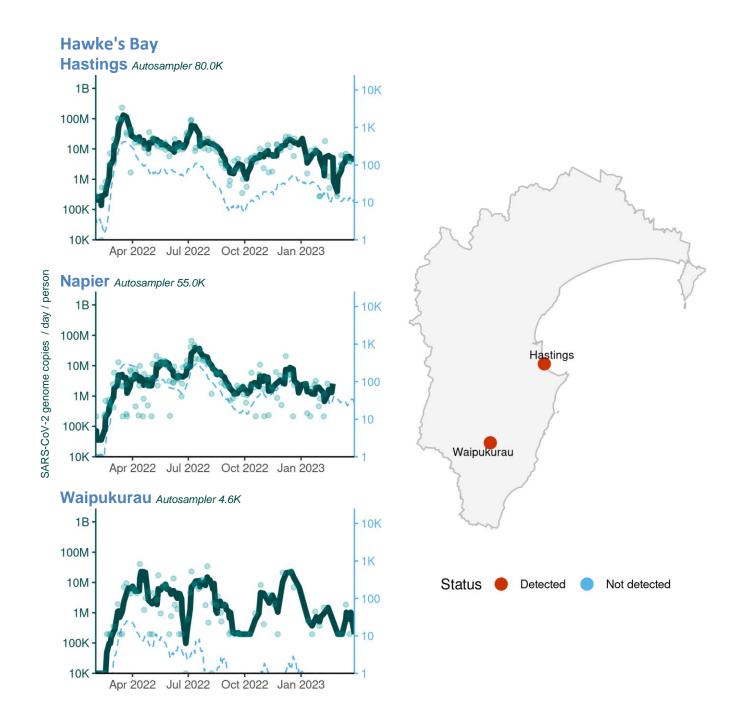


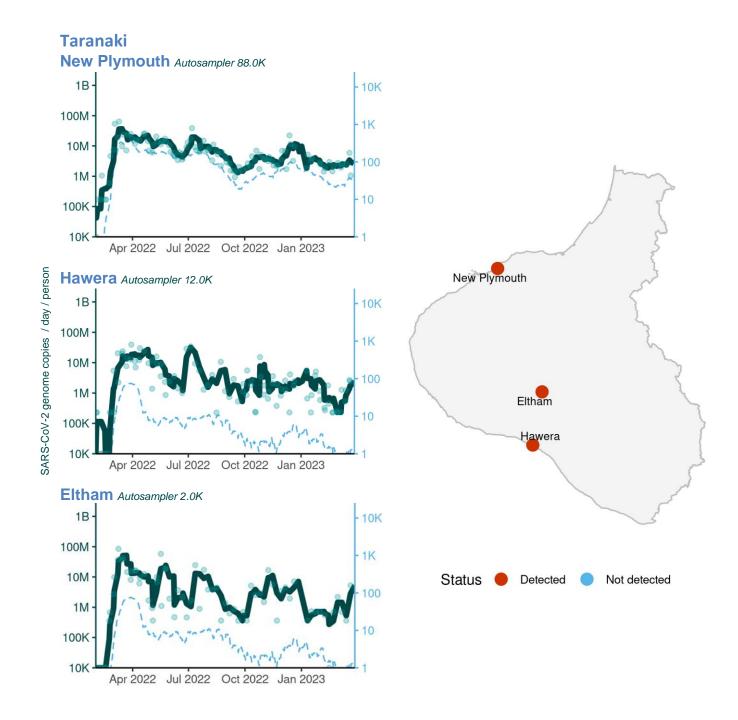


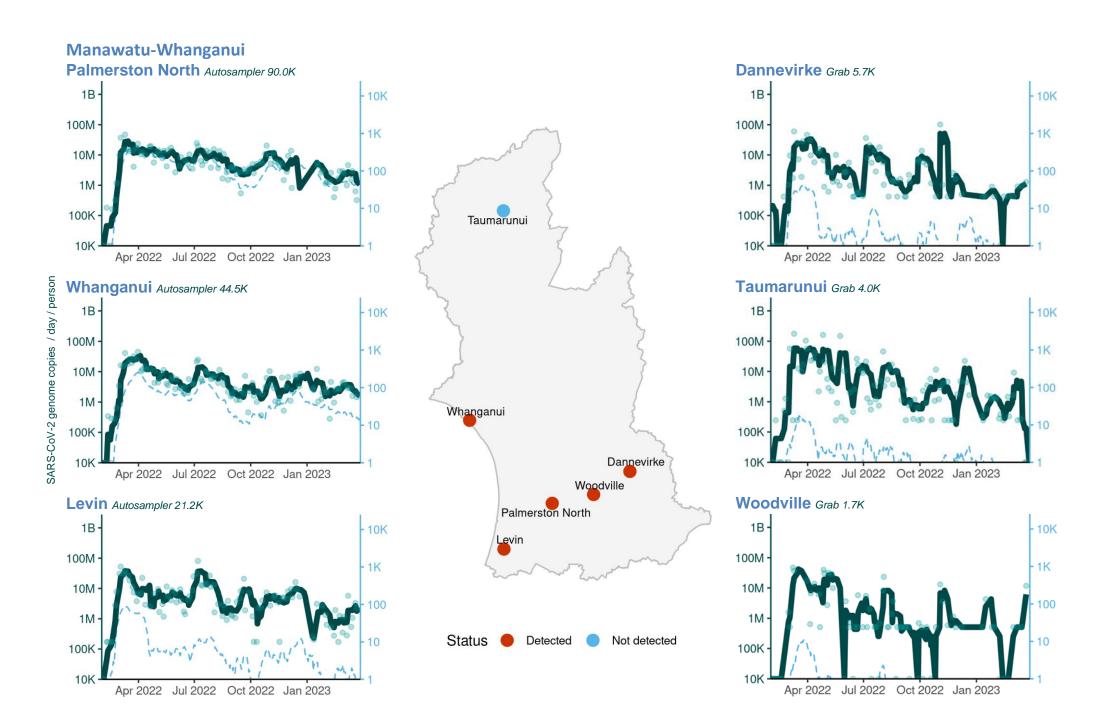


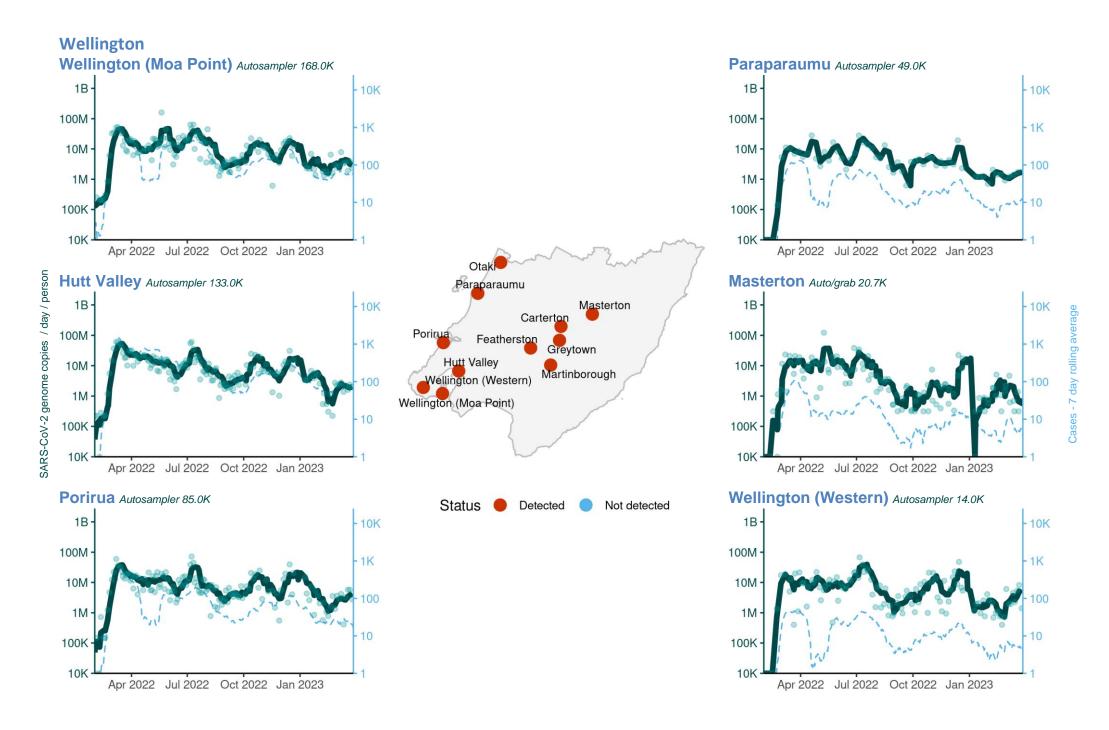


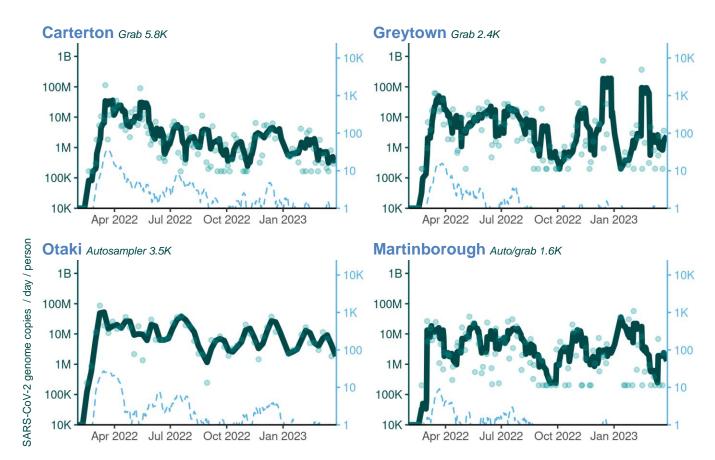


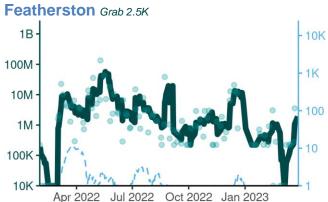


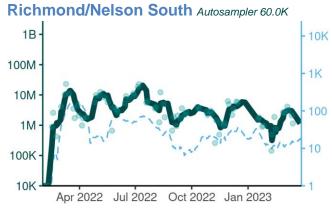




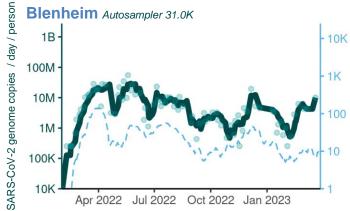


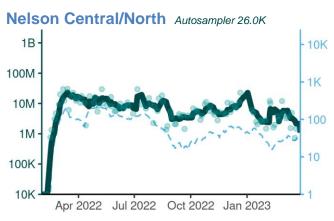




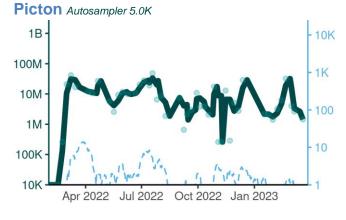


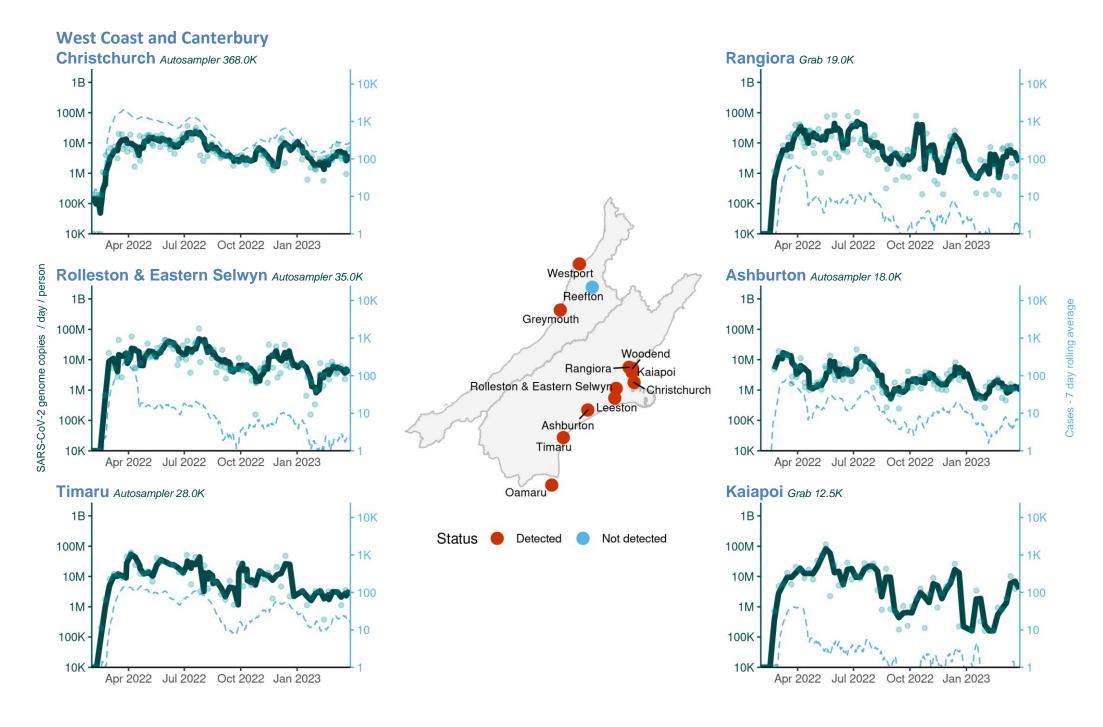
Tasman, Nelson, and Marlborough

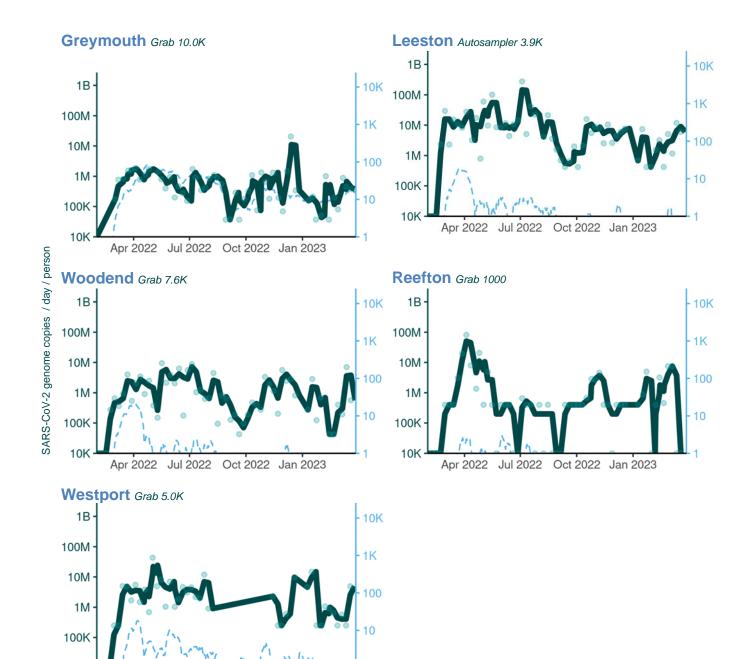






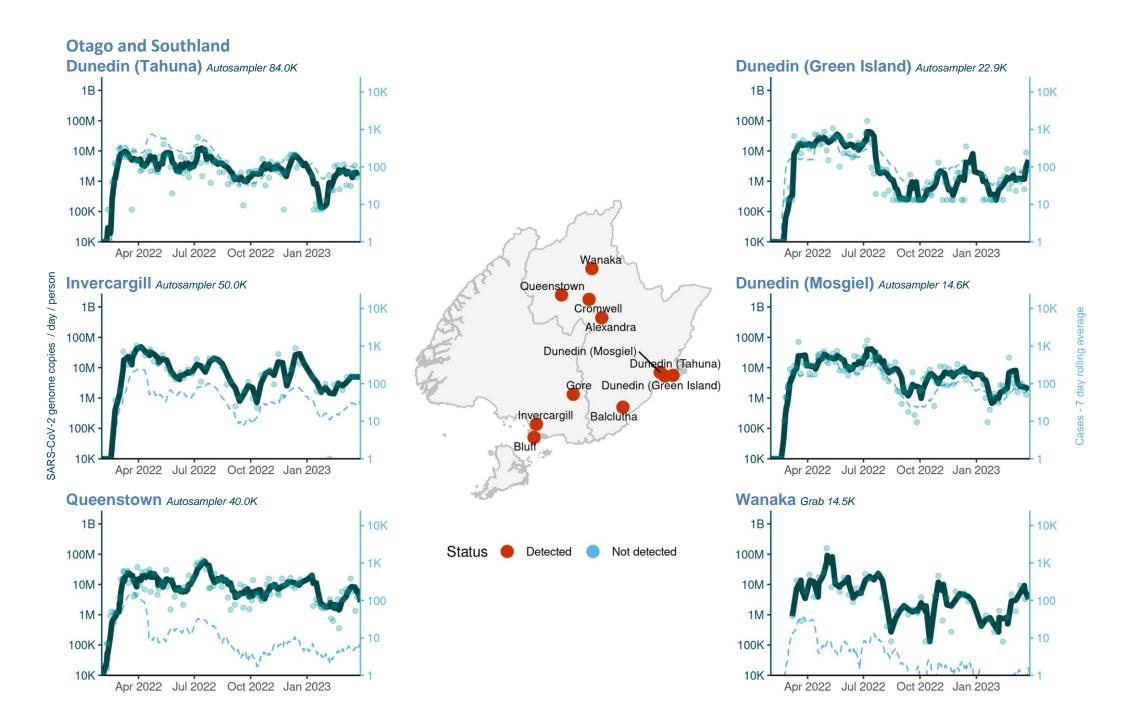






10K

Apr 2022 Jul 2022 Oct 2022 Jan 2023



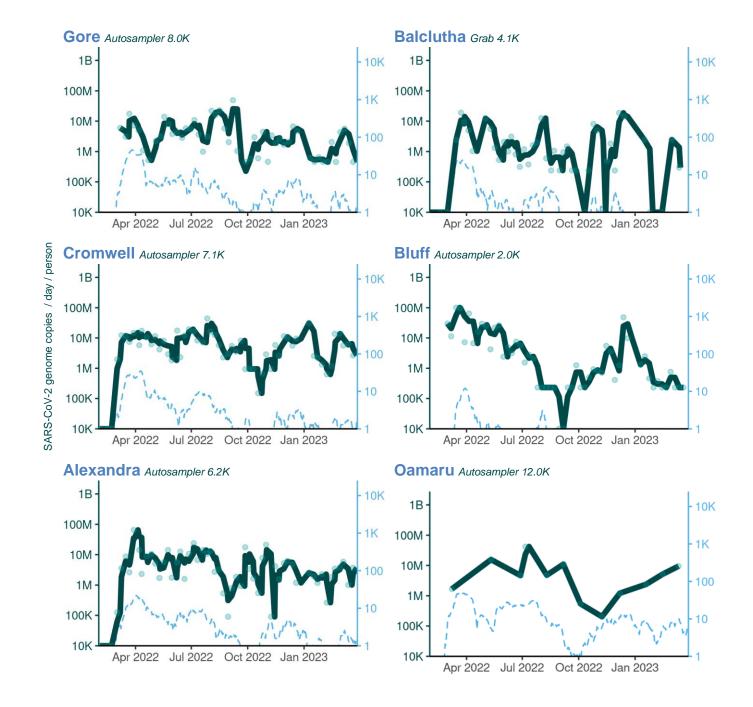


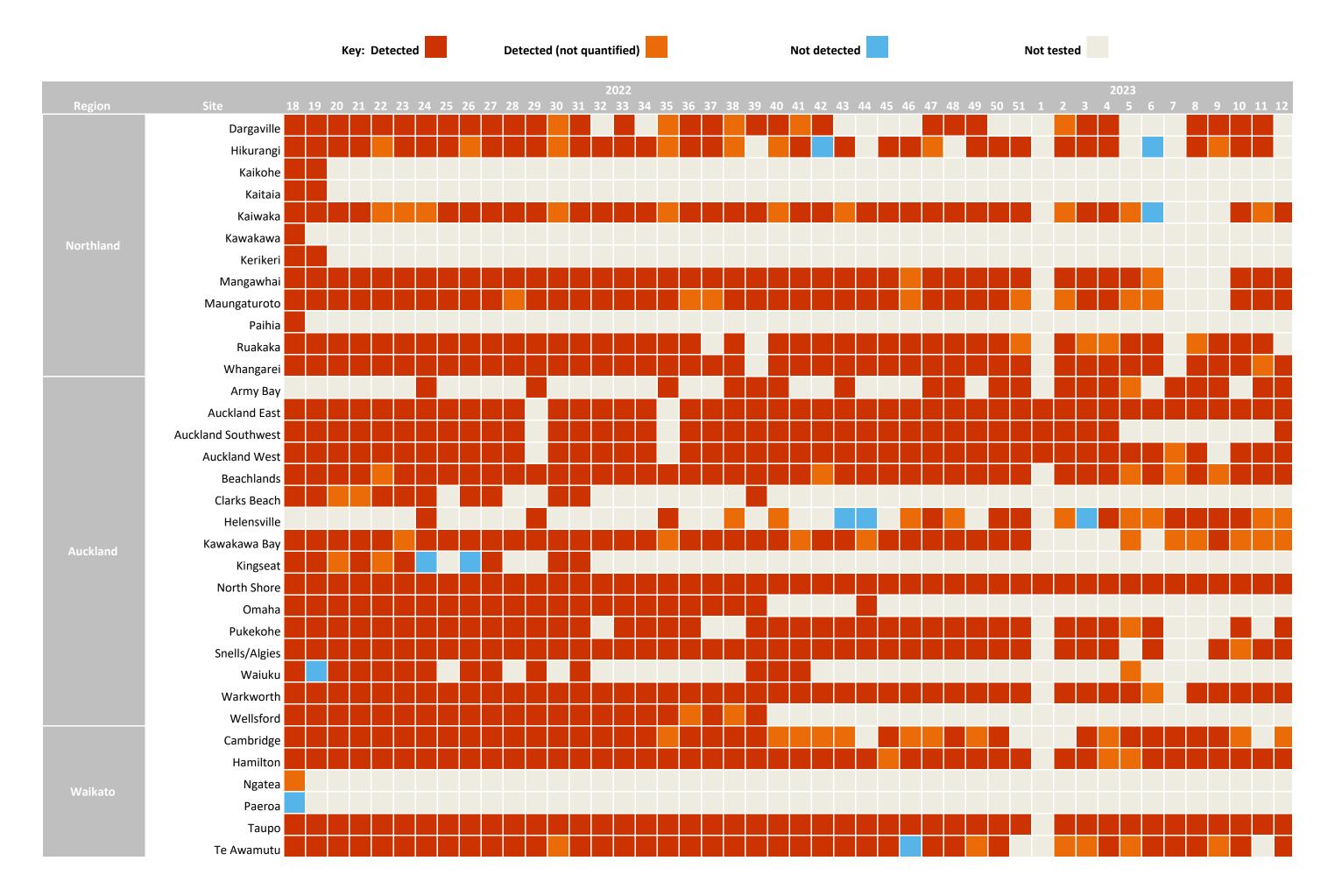
Table 2: Results for weeks 11& 12(ending 19 March and 26 March 2023)

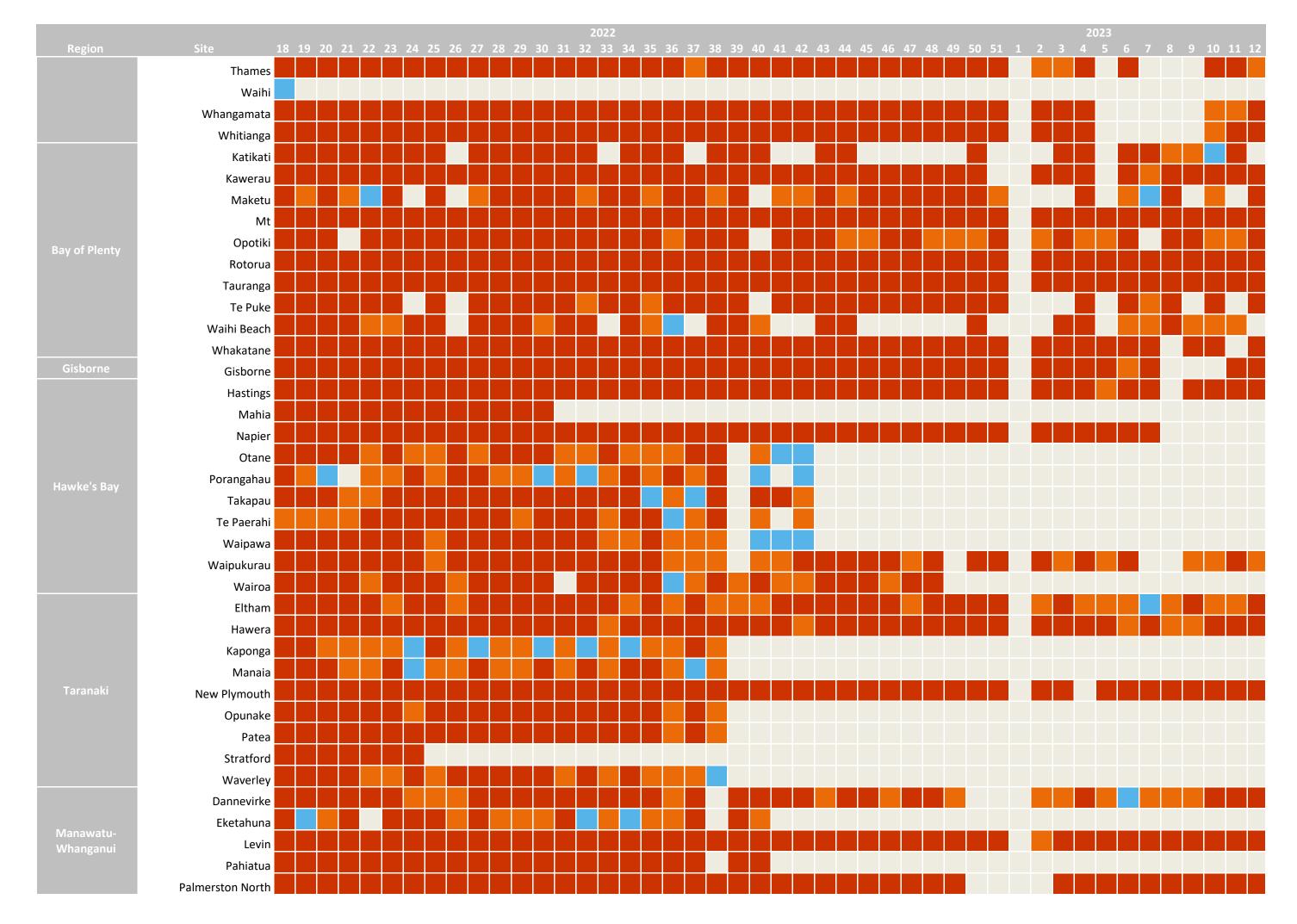
Wastewater testing results. Grab samples are collected usually over 15-30 minutes. Autosampler are 24-hour composites.

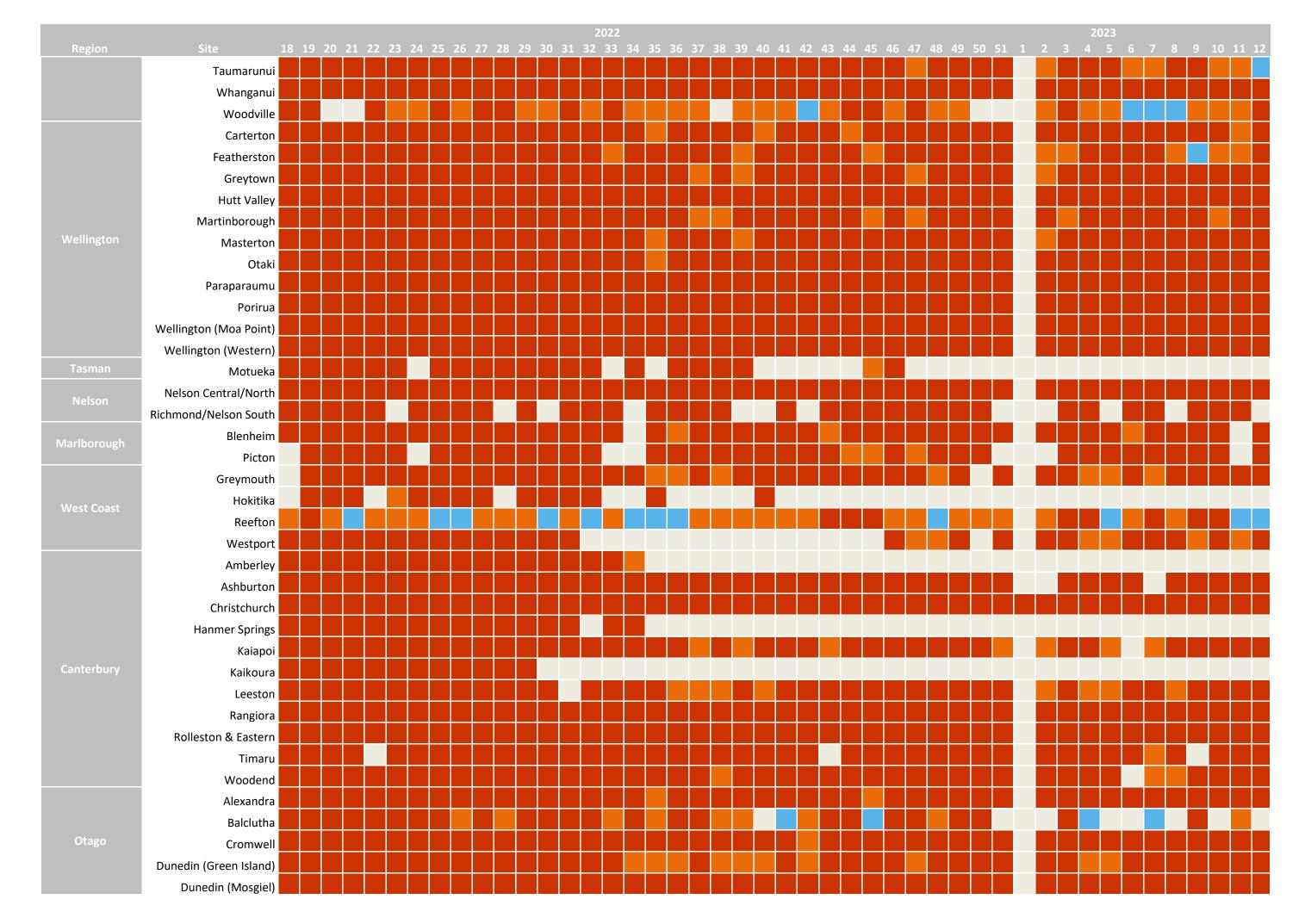
									March 2023					
Region	Site	Population	Sample Type	Mon 13	Tue 14	Wed 15	Thu 16	Mon 20	Tue 21	Wed 22	Thu 23	Fri 24	Sat 25	Sun 26
Northland	Dargaville	5,000	Grab				Detected							
	Hikurangi	1,730	Grab		Detected									
	Kaiwaka	400	Grab	Detected			Detected	Detected			Detected			
	Mangawhai	1,100	Grab	Detected			Detected	Detected			Detected			
	Maungaturoto	1,300	Grab	Detected			Detected	Detected			Detected			
	Ruakaka	4,500	Grab			Detected								
	Whangarei	65,000	Autosampler			Detected							Detected	Detected
Auckland	Army Bay	42,000	Autosampler		Detected				Detected					
	Auckland East	680,000	Autosampler	Detected		Detected		Detected		Detected				
	Auckland Southwest	120,000	Autosampler					Detected		Detected				
	Auckland West	315,000	Autosampler	Detected		Detected		Detected		Detected				
	Beachlands	6,760	Grab			Detected				Detected				
	Helensville	3,800	Autosampler		Detected				Detected					
	Kawakawa Bay	600	Grab			Detected				Detected				
	North Shore	240,000	Autosampler	Detected		Detected		Detected		Detected				
	Pukekohe	20,900	Autosampler							Detected				
	Snells/Algies	4,000	Autosampler		Detected				Detected					
	Warkworth	3,500	Autosampler		Detected				Detected					
Waikato	Cambridge	20,100	Autosampler							Detected				
	Hamilton	169,000	Autosampler	Detected	Detected			Detected						
	Taupo	23,000	Auto/grab		Detected		Detected		Detected		Detected			
	Te Awamutu	13,100	Autosampler							Detected				
	Thames	7,500	Autosampler			Detected					Detected			
	Whangamata	4,000	Autosampler			Detected					Detected			
	Whitianga	6,600	Autosampler			Detected					Detected			
Bay of Plenty	Katikati	5,500	Autosampler			Detected								
	Kawerau	7,000	Autosampler		Detected				Detected					
	Maketu	1,300	Autosampler							Detected				
	Mt Maunganui/Papamoa		Autosampler		Detected		Detected		Detected		Detected			
	Opotiki	-	Autosampler		Detected		Detected		Detected		Detected			
	Rotorua	59,000	Autosampler		Detected		Detected		Detected		Detected			
	Tauranga	50,000	Autosampler		Detected		Detected		Detected		Detected			
	Te Puke	9,700	Autosampler							Detected				
	Waihi Beach	3,600	Autosampler			Detected								
	Whakatane	21,020	Autosampler						Detected					
Gisborne	Gisborne	37,000	Autosampler	Detected				Detected		Detected				
Hawke's Bay	Hastings	80,000	Autosampler		Detected	Detected			Detected	Detected				
	Waipukurau	4,610	Autosampler			Detected			Detected					
Taranaki	Eltham	2,006	Autosampler			Detected				Detected				
	Hawera	12,000	Autosampler		Detected	Detected			Detected	Detected				
	New Plymouth	88,000	Autosampler				Detected		Detected		Detected			
Manawatu-	Dannevirke	5,696					Detected				Detected			
Whanganui	Levin	21,200	Autosampler		Detected		Detected		Detected		Detected			
	Palmerston North	90,000	Autosampler		Detected		Detected		Detected		Detected			
	Taumarunui	4,000	Grab		Detected				Not detected		Not detected			
	Whanganui		Autosampler	Detected			Detected		Detected		Detected			
	Woodville	1,657	Grab				Detected				Detected			

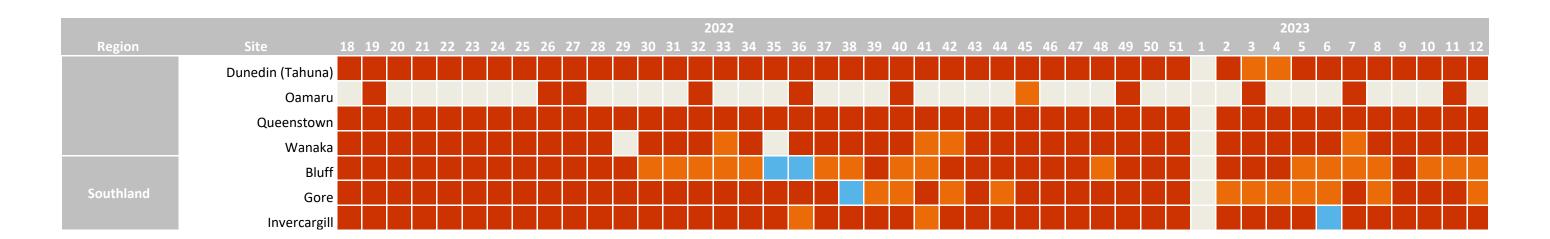
									March 2023					
Region	Site	Population	Sample Type	Mon 13	Tue 14	Wed 15	Thu 16	Mon 20	Tue 21	Wed 22	Thu 23	Fri 24	Sat 25	Sun 26
Wellington	Carterton	5,800	Grab	Detected			Detected	Detected				Detected		
	Featherston	2,500	Grab		Detected		Detected		Detected		Detected			
	Greytown	2,438	Grab		Detected		Detected		Detected		Detected			
	Hutt Valley	133,000	Autosampler	Detected			Detected	Detected			Detected			
	Martinborough	1,641	Auto/grab		Detected		Detected		Detected		Detected			
	Masterton	20,700	Auto/grab		Detected	Detected			Detected	Detected				
	Otaki	3,500	Autosampler		Detected				Detected					
	Paraparaumu	49,000	Autosampler		Detected				Detected					
	Porirua	85,000	Autosampler	Detected			Detected	Detected			Detected			
	Wellington (Moa Point)	168,000	Autosampler	Detected			Detected	Detected			Detected			
	Wellington (Western)	14,000	Autosampler	Detected			Detected	Detected			Detected			
Nelson	Nelson Central/North	26,000	Autosampler	Detected		Detected		Detected		Detected				
	Richmond/Nelson South	60,000	Autosampler	Detected		Detected								
Marlborough	Blenheim	31,000	Autosampler						Detected					
	Picton	5,000	Autosampler						Detected					
West Coast	Greymouth	10,000	Grab	Detected						Detected				
	Reefton	1,000	Grab	Not detected				Not detected						
	Westport	5,000	Grab	Detected				Detected						
Canterbury	Ashburton	18,000	Autosampler		Detected	Detected		Detected						
	Christchurch	368,000	Autosampler	Detected		Detected		Detected		Detected				
	Kaiapoi	12,500	Grab		Detected				Detected					
	Leeston	3,900	Autosampler	Detected				Detected						
	Rangiora	19,000	Grab		Detected		Detected		Detected		Detected			
	Rolleston & Eastern Selwyn	35,000	Autosampler	Detected				Detected		Detected				
	Timaru	28,000	Autosampler		Detected				Detected					
	Woodend	7,600	Grab		Detected				Detected					
Otago	Alexandra	6,200	Autosampler		Detected					Detected				
	Balclutha	4,100	Grab		Detected									
	Cromwell	7,100	Autosampler		Detected					Detected				
	Dunedin (Green Island)	22,900	Autosampler	Detected			Detected	Detected			Detected			
	Dunedin (Mosgiel)	14,600	Autosampler	Detected			Detected	Detected			Detected			
	Dunedin (Tahuna)	84,000	Autosampler	Detected			Detected	Detected			Detected			
	Oamaru	12,000	Autosampler	Detected										
	Queenstown	40,000	Autosampler	Detected		Detected			Detected		Detected			
	Wanaka	14,500	Grab	Detected					Detected					
Southland	Bluff	2,000	Autosampler		Detected				Detected					
	Gore	8,000	Autosampler			Detected				Detected				
	Invercargill	50,000	Autosampler		Detected				Detected					

Table 3: Weekly Summary of Wastewater Sampling Results for SARS-CoV-2









#### **Acknowledgements**

This work represents the combined efforts of a large number of individuals and organisations.

We continue to be indebted to the teams across the country who are collecting the wastewater that underpins this work.

The wastewater analysis has been undertaken at ESR by a team which may on any given week include contributions from Joanne Chapman, Dawn Croucher, Joanne Hewitt, Joycelyn Ho, Anower Jabed, Ashley McDonald, Andrew Ng, and Fatiha Sulthana. Data science analysis, visualisation and reporting is the result of team effort from: Franco Andrews, Bridget Armstrong, Raewyn Campbell, Joanne Chapman, Lei Chen, Gerhard de Beer, Richard Dean, Brent Gilpin, Joanne Hewitt, Dawen Li, Jonathan Marshall, Helen Morris and Leighton Watson. Ongoing support for this work from the Ministry of Health and ESR management is appreciated.

#### **Notes**

Sites and frequency of sample collection: The catchment population sites selected for the surveillance range from approximately 400 to over 1,000,000 individuals. The sites cover all regions of the country. Most major towns and all cities, as well as many smaller communities, are included. In early 2023, the wastewater catchment areas cover over 75% of the population connected to wastewater treatment plants. The sites from which samples have been collected have varied over the last 12 months. New sites may be added over time, and/or sampling may reduce in frequency or cease for other sites. The selection and frequency of sampling vary depending on the local population, access to wastewater collection points, staff availability to collect samples and risk factors. When included, samples are collected at least weekly, with twice weekly sampling being common.

Sampling method: The preferred option is to automatically collect a 24 hour 'composite' sample. This is where a pump automatically collects a small volume of wastewater every 15 minutes over 24 hours using a composite sampler. These samplers are available in some wastewater treatment plants. When composite samplers are not available, 'grab' samples are collected. These range from a sample being taken at a single point in time, to 3 samples taken over 30 minutes, to samples collected over a day. Grab samples represent only the composition of the source at that time of collection and may not be as representative as a 24-hour composite sampler. More variation may be expected with grab samples.

Laboratory analysis of wastewater samples: Samples are sent from each wastewater treatment plant to ESR. Processing of each sample commences within an hour or two of receipt. Processing involves the concentration of virus from 250 mL sample to approx. 1 mL using centrifugation and polyethylene glycol. Viral RNA is then extracted from a small volume of 0.2 mL concentrate to give a final volume of 0.05 mL The presence of SARS-CoV-2 RNA is determined using RT-qPCR. SARS-CoV-2 is considered detected when any of the RT-qPCR replicates are positive.

**RT-qPCR:** Reverse transcription (RT) to convert RNA to complementary DNA (cDNA), followed by quantitative PCR (qPCR). RT-qPCR is used for detection and quantification of viral RNA.

**Method sensitivity:** The protocol used to concentrate SARS-CoV-2 from wastewater allows for the sensitive detection of SARS-CoV-2 by RT-qPCR. ESR has shown that when 10 individuals are actively shedding SARS-CoV-2 RNA in a catchment of 100,000 individuals, there was a high likelihood of detecting viral RNA in wastewater (https://doi.org/10.1016/j.watres.2021.118032). Shedding by one individual may be detected in wastewater, but it does depend on many factors including the amount and duration of shedding. Very low levels in wastewater may be not able to be quantified (i.e., less than the limit of quantification - see below).

SARS-CoV-2 RNA detected (positive result): A positive detection in the wastewater indicates that at least one person has been shedding SARS-CoV-2 into the wastewater at some point during the time period that the sample was being collected. In some cases, detections could also be due to the shedding of low levels of SARS-CoV-2 RNA by a recently recovered case. The detection of SARS-CoV-2 RNA does not indicate that infectious virus is present.

**SARS-CoV-2 RNA not detected (negative result):** A negative result can occur because there are no active 'shedding' cases in the catchment or because the SARS-CoV-2 RNA concentration is too low to be detected, most likely because there are a very low number of cases in the wastewater catchment. Therefore, negative finding does not necessarily guarantee the absence of COVID-19 in the community.

Viral loads and normalisation: When detected, the SARS-CoV-2 RNA concentration is calculated as genome copies per L of wastewater. This is then converted to a viral load of genome copies/day/person. This conversion takes into account the flow rate of wastewater entering the treatment plant (the influent) and the population in the catchment. The flow rate is the total volume (m3 per day) recorded at the inlet of the wastewater treatment plant over 24 hours. This is a population-normalised viral load. Currently, the flow rate is the average annual flow rate, but will be replaced with daily flow rate when available (note that rainfall may significantly increase the flow rate at the inlet, diluting the sample, and may result in lower concentrations and a false negative result).

**Limit of quantification:** The lowest concentration of the target that can be reliably quantified is referred to as the limit of quantification. For those samples where SARS-CoV-2 is detected but cannot be quantified, a value of 5 genome copies/mL wastewater is used. While a standard method is being used, virus recovery can vary from sample to sample, and this may affect the quantitation.

**Data subject to change:** Data generated for the New Zealand Wastewater COVID-19 Surveillance Programme should be considered provisional and may be subject to change. Data may be incomplete for the most recent 2-week period due to processing, testing and reporting delays.

#### Data not shown:

• Results from certain samples may not be shown, as the result was either deemed invalid, or the sample could not be tested (e.g., leaked in transit, not labelled).

#### For further information please contact:

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